Slater E C & Bonner W D. The effect of fluoride on the succinic oxidase system.
Biochemical J. 52:185-96, 1952.
[Molteno Institute, University of Cambridge, England]

Spectrophotometric methods for measuring succinate oxidase and dehydrogenase are described. Fluoride and phosphate separately, but more strongly when acting together, are competitive inhibitors of succinate dehydrogenase. The limiting assumptions of the Michaelis-Menten equation are not applicable to this enzyme; the Briggs-Haldane equation should be used. A method of determining the rate constants for association of substrate and enzyme and dissociation of the enzyme-substrate complex is given. [The *SCI®* indicates that this paper has been cited in over 570 publications.]

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Fluoride and Succinate Dehydrogenase Kinetics

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E.C. Slater
Department of Biochemistry
University of Southampton
Bassett Crescent East
Southampton SO9 3TU
England
and
Walter D. Bonner
Department of Biochemistry and
Biophysics
School of Medicine
University of Pennsylvania
Philadelphia, PA 19104-6089

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This is the last of a series of papers on the succinate oxidase system, starting in 1949,[1] that one of us (ECS) published in the *Biochemical Journal* while working under the direction of David Keilin. By 1950 his interests had turned to the mechanism of oxidative phosphorylation and when a postdoctoral visitor (WDB) to the institute found that phosphate is necessary for maximal inhibition of succinate oxidation[2] by fluoride, we joined forces, in the hope that a further investigation of this requirement might provide a clue towards the mechanism of oxidative phosphorylation.

It did not. However, it did resolve conflicting conclusions in the literature concerning the locus of action of fluoride on respiratory systems. We were able to show that either fluoride or phosphate alone acts as a classical competitive inhibitor of succinate de-

hydrogenase. The two together are, however, much more effective, suggesting the formation of fluorophosphate in the active site of the enzyme. Using the classical procedures of enzyme kinetics, we measured the various inhibitory constants. However, there remained a problem. We confirmed a previous finding by Massart that the degree of inhibition is greater when the complete succinate oxidase system is measured than when its component, succinate dehydrogenase, is determined with methylene blue as acceptor.

The degree of inhibition is determined by the ratio K_i/K_m, where K_i is the dissociation constant of the inhibitor-enzyme complex and K_m is the Michaelis constant of the enzyme. It was widely believed at that time that K_m is identical with the dissociation constant of the enzyme-substrate complex, which would be independent of the hydrogen acceptor. However, already in 1925, Briggs and Haldane had shown that a more general expression for K_m is $(k_2 + k_3)/k_1$, where the reaction constants are defined by

$$E + S \underset{k_2}{\overset{k_1}{\rightleftarrows}} ES \overset{k_3}{\rightarrow} E + P.$$ This reduces to $k_2/k_1 = K_D$,
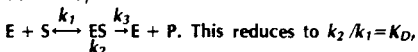
when $k_2 > k_3$. It occurred to us that, if the Briggs-Haldane equation were applicable to succinate dehydrogenase, then, since k_3 and therefore K_m would be dependent on the hydrogen acceptor and its concentration, the corresponding dependence of the degree of inhibition by a competitive inhibitor would be explained. This idea was confirmed by the demonstration of a linear relationship between V (the velocity at infinite substrate concentration)/v (the observed velocity) at different values of V, obtained by varying the nature of the acceptor and its concentration. By extrapolation of this straight line to $V = O$, k_2 could be calculated. This demonstration of the validity of the Briggs-Haldane equation we considered to be the most important part of our paper.

The paper and a more general exposition[3] during a symposium on enzymes organized by the Faraday Society aroused considerably more interest than we had expected, and we had the satisfaction of seeing the method for determining k_2 included in M. Dixon and E.C. Webb's book.[4]

However, this is now all old hat and cannot be the reason for the high citation score. Perhaps, the interest in fluoride toxicology and pharmacology is a partial explanation. However, consultation of some recent citations has probably revealed the main reason. We had completely forgotten that, in this paper, we had introduced convenient spectrophotometric methods for measuring succinate dehydrogenase and succinate oxidase. It appears that these methods are still widely used for measuring the activity of this enzyme as a mitochondrial marker (see, for example, reference 5).

1. Slater E C. A comparative study of the succinic dehydrogenase-cytochrome system in heart muscle and in kidney. *Biochemical J.* 45:1-8, 1949. (Cited 250 times.)
2. Bonner W D. Activation of the heart-muscle succinic oxidase system. *Biochemical J.* 49:viii, 1951. (Cited 15 times.)
3. Slater E C. Calculation of the rate constants of the reaction between an enzyme and its substrate from the overall kinetics of the reaction catalysed by the enzyme. *Discuss. Faraday Soc.* 20:231-40, 1955. (Cited 45 times.)
4. Dixon M & Webb E C. *Enzymes.* New York: Academic Press, 1958. 782 p. (Cited 565 times.)
5. Takeo S, Miyake K, Minematsu R, Tanonaka K & Konishi M. *In vitro* effect of naftidrofuryl oxalate on cerebral mitochondria impaired by microsphere-induced embolism in rats. *J. Pharmacol. Exp. Ther.* 248:1207-14, 1989.

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